Synthesis and Anti-HIV Evaluation of Some 5'-0-Phosphonomethyl-2',3'-Dideoxynucleosides

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5'-0-Phosphonomethylation of different pyrimidine 2',3'-dideoxyribosides was accomplished by reaction of the latter with diethyl p-toluenesulfonyloxymethane phosphonate in the presence of sodium hydride. The basephosphonylated and sugar-phosphonylated derivatives could be readily distinguished by spectroscopic techniques. Protection of the uracil or thymine moiety with a N3-benzoyl group failed to prevent base modification. However, 04-methyl-protected 2',3'-dideoxyuridine readily afforded the 5'-O-phosphonylated derivative, which was then converted to both the 2',3'-dideoxyuridine and 2',3'-dideoxycytidine analogues. The 5'-O-phosphonomethyl derivatives of 3'-deoxythymidine, 2',3'-dideoxyuridine, 2',3'-dideoxycytidine, 3'-O-methylthymidine and 3'-amino-3'-deoxythymidine did not show an appreciable anti-HIV activity in MT-4 cells. In contrast, the 5'-O-phosphonomethyl derivatives of 3'-fluoro-3'-deoxythymidine and 3'-azido-3'-deoxythymidine achieved 50% inhibition of HIV-1 cytopathogenicity at a concentration of approximately 1 µM, which is 1000-fold higher than the concentration at which 3'-fluoro-3'-deoxythymidine inhibits HIV-1 replication in MT-4 cells. Also, the 5'-0-diphosphoryl derivative of 5'-0-phosphonomethyl-3'-deoxy-3'-fluorothymidine showed a 1000-fold lower affinity for HIV-1 reverse transcriptase than the 5'-triphosphate of 3'-deoxy-3'-fluorothymidine.

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Anti-HIV-1 Activity of Antiviral Compounds, as Measured by a Quantitative Focal Iramunoassay in CD4<sup>†</sup> HeLa Cells and a Plaque Assay in MT-4 Cells H. Nakashima, D. Schols, R. Pauwels, J. Balzarini, J. Desmyter and E. De

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Focus- and plaque-forming assays are often used in virological research to quantitate virus in biological samples. These rechniques are known for their efficiency, simplicity, reproducibility, and reliability. The antihuman immunodeficiency virus type 1 (HIV-1) activities of selected nucleoside analogues and sulfated polysaccharides were evaluated by a plaque assay (PA) in MT-4 cells and a focal immunoassay (FIA) in CD4 HeLa cells. Similar 50% inhibitory concentrations (IC50) were obtained for the sulfated polysaccharides when measured by PA or FIA; the IC50 values of dextran sulfate and pentosan polysulfate were 0.80 µg/ml and 0.35 µg/ml, respectively. Also, comparable IC50 values were obtained for the purine 2',3'-dideoxyribosides DDA, DDI and DDG, when evaluated by PA or FIA: their IC50 values ranged from 1.42 to 2.71 µM. In contrast, the IC50 values of the pyrimidine 2'.3'-dideoxyribosides were invariably 4- to 10-fold lower when monitored by PA than if measured by FIA: IC50 of AZT, D4T and DDC, as based on PA, were 0.015 µM, 0.094 µM and 0.038 µM, respectively; their IC50 values, as based on FIA, were 0.062 µM, 0.285 µM and 0.463 µM, respectively. The differences in anti-HIV-1 activity found for AZT, D4T and DDC when evaluated in the PA and FIA assay systems may at least in part be related to differences in the metabolism (i.e. thymidine kinase and/or 2'-deoxycytidine kinase levels) between MT-4 and CD4 HeLa cells.